LAB SAFETY TRAINING

All new and returning students/visitors/volunteers are required to take the annual lab safety training before working in any lab. Lab Safety Training is now available through the UR Blackboard System—Directions are below.

Access to Blackboard for students is best done on a UR computer. Lab safety training needs to be done within a week of the student arrival and/or working in a lab.

DIRECTIONS FOR STUDENTS/VOLUNTEERS TO ACCESS BLACKBOARD

Access the UR intranet. In the search box, type “Blackboard.”

On the Listings page – (see Blackboard Support, Edward G. Miner Library-Rochester).

Click on that listing where there will be a question, “Request an Account”.

Once clicked – select “Don’t have an account – create a basic account”.

This will take them into the Blackboard system to take the course training.

Any difficulties with access – they need to call the Blackboard Helpdesk at 5-6865.

NEW STUDENTS/NEW YEAR
LAB SAFETY REVIEW

It’s time to return to the classroom. Let’s review some safety precautions for NEW student/staff arrivals in the laboratory.

DRESS FOR SUCCESS
Success in completing your experiments safely. The University’s Chemical Safety Program, which applies to all labs where chemicals are present, states the following: (http://www.safety.rochester.edu/ih/chp/chpplan-15.html):

Personal Apparel
Severe injuries and death have been reported to have occurred to lab personnel because their hair was caught in moving lab equipment or inappropriate clothing was worn. Although the PI/supervisor is responsible for having his/her personnel wear appropriate apparel, EH&S requires lab personnel:

- to confine or tie long hair and loose clothing;
- not wear clothing that contains dangling laces/strings, including neckties;
- not wear dangling jewelry;
- utilize break-away lanyards for IDs;
- wear only closed toed shoes/sneakers;
- never wear high-heeled shoes or light weight shoes/slippers that do not provide protection from broken glass or hazardous materials that may be on the floor and are not suitable for the work environment;
- wear lab coats over street clothes to minimize potential fire hazards or chemical contamination when working with chemicals, especially flammable and pyrophoric materials;
- utilize natural fibers (cotton) when working with flammable materials as this material is less combustible than synthetic fabrics; wear eye and face protection; and, wear hearing protection when required.
- Failure to adhere to these minimum requirements has lead to many laboratory injuries and exposures, here at the University and across the country in other academic labs. Be Proactive—DRESS FOR SUCCESS!

LAB SAFETY MANUAL
- Each laboratory is equipped with a lab safety manual that is updated annually. If you have any questions about specific lab practices—please seek out your lab manager—he or she can give you all the information necessary to maintain a safe lab environment at all times.
- Not sure ?? We love questions and we will find you an answer—call the IBC office at 5-2402.

****IMPORTANT INFORMATION*****

Biosafety Cabinet Certification and Repair

The U of R has introduced a new contractor for your Biosafety Cabinet needs. Their rates are very reasonable.

Name: B & V Testing
Blanket Order #: UK730429-14
For: 7/1/13-6/30/14

Contact: Sandy MacLean-Horne
phone: 800.851.9081 ext. 103
fax: 781.647.3770

Link to their website: http://www.bandvtesting.com/technical-services/biosafety-cabinet-certification/
# Good Microbiological Practices

## Procedures to Minimize Aerosol Hazards

### Opening Tubes
- Manipulate infectious materials within a biological safety cabinet.
- Upon opening, unscrew the cap slightly and wait a few seconds before removing it.

### Pipetting
- Use “to deliver” pipettes calibrated to retain the last drop.
- Use pipettes with plugs.
- Discharge pipettes close to the fluid level and let the contents run down the wall of the container.
- Never forcefully expel infectious materials from the pipette.

### Breakage
- Avoid the use of glassware where possible.
- Use plastic tubes, flasks and bottles.
- Use screw-capped tubes and bottles rather than plugs or snap caps.

### Inoculating Loop
- Use a micro-inoculator or a disposable loop instead of a bunsen burner.
- Allow the inoculating loop to cool before any procedures.

### Centrifugation
- Centrifuge infectious material in closed containers, placed in sealed safety cups or rotors.
- Open cups in a biological safety cabinet.
- Maintain the centrifuge to ensure that it is clean and the gaskets and O-rings are not compromised.
- Wait 5 minutes before opening the centrifuge after each run to allow any aerosols to settle.

### Mixing and Homogenizing
- Ensure the lab blender has a gasket lid and leak proof bearings.
- Wait a few seconds before opening a lid after mixing.
- Use a vortex, instead of inverting the cultures.

### Syringes/Needles
- Withdraw needles from bottles using disinfectant-soaked absorbent pads wrapped around the bottle cap.
- Use locking syringes.

### Pouring Infectious Materials
- Perform your work over plastic-backed absorbent material.
- Wipe the rim of the tube with disinfectant-soaked absorbent paper to remove potential contamination on the outside of the tube.